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# WO<sub>3</sub> photocatalyst containing copper inactivates SARS-CoV-2 Pango lineage A and Omicron BA.2 variant in visible light and in darkness

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Abstract: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the causative agent of 19 coronavirus disease 2019, which has been a global pandemic. Since SARS-CoV-2 is transmitted 20 through contaminated surfaces and aerosols, environmental disinfection is important to block the 21 spread of virus. Photocatalysts are attractive tools for virus inactivation and are widely used as air 22 purifiers and coating materials. However, photocatalysts are inactive in the dark and some of them 23 need to be excited with a light of specific wavelength. Therefore, photocatalysts that can effectively 24 inactivate SARS-CoV-2 in indoor environments are needed. Here we show that a WO<sub>3</sub> photocatalyst 25 containing copper inactivated the SARS-CoV-2 WK-521 strain (Pango lineage A) upon irradiation 26 with white light, in a time- and concentration-dependent manner. Additionally, this photocatalyst 27 also inactivated SARS-CoV-2 in dark condition, due to the antiviral effect of copper. Furthermore, 28 this photocatalyst inactivated not only the WK-521 strain but also the Omicron variant BA. 2. These 29 results indicate that the WO<sub>3</sub> photocatalyst containing copper can inactivate indoor SARS-CoV-2 30 regardless of the variant, in visible light or darkness, making it an effective tool for controlling the 31 spread of SARS-CoV-2. 32

Keywords: SARS-CoV-2 inactivation; Pango lineage A; Omicron variant BA. 2; WO3 photocatalyst;33time-dependency; dose-dependency; copper based disinfection; environmental disinfection34

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# 1. Introduction

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the causative 37 agent of coronavirus disease 2019 (COVID-19), which has had an unprecedented impact 38 on modern human civilization [1], and resulted in more than 6.3 million deaths globally, 39 as of early June 2022. Despite the development of drugs and vaccines, the number of in-40 fected people continues to increase. Although the transmission route of SARS-CoV-2 is 41 still being debated, it is generally believed to be transmitted through the airborne route, 42 surface contamination, and fecal-oral transmission [2]. Thus, inactivation of the virus in 43 the air and on surfaces is essential for controlling its transmission. In addition, the genome 44 of SARS-CoV-2 has mutated rapidly, and several variants are reported. Mutations in virus 45

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**Copyright:** © 2022 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/). help it to evade the host immune system and to acquire drug resistance. Therefore, despite
the presence of vaccines and drugs, it is important to find effective ways to inactivate the
virus to prevent the spread of infection, regardless of the variant. It is reported that SARSCoV-2 can be inactivated by photocatalysts [3], heat [4], ultraviolet (UV) [5,6] and disinfectants such as ethanol [7]. Especially, since photocatalysts are harmless to human body
unlike UV, they have recently received great attention and it is proposed that they can be
applied for disinfection of living and working spaces, without evacuating people.

Photocatalysts are excited by light and exhibit a strong oxidation-reduction reaction 53 generating reactive oxygen species (ROS), such as hydroxyl ('OH) and superoxide radicals 54 (O<sub>2</sub>-), on their surface [8]. Using this oxidation-reduction reaction, photocatalysts kill mi-55 croorganisms, such as bacteria and fungi, and inactivate viruses such as influenza virus, 56 hepatitis C virus, vesicular stomatitis virus, enterovirus, herpes virus, Zika virus, human 57 coronavirus, bovine coronavirus, human norovirus, murine norovirus, SARS coronavirus, 58 and bacteriophage [8-15]. Many compounds such as titanium dioxide (TiO<sub>2</sub>), tungsten 59 trioxide (WO<sub>3</sub>), zinc oxide (ZnO), cadmium sulfide (CdS), and iron (III) oxide (Fe<sub>2</sub>O<sub>3</sub>) are 60 known to exhibit photocatalysis and are being actively researched. In particular, TiO<sub>2</sub> and 61 WO<sub>3</sub> have been reported to inactivate SARS-CoV-2, and are very promising as antiviral 62 materials [3,16,17]. In addition, photocatalysts damage the viral morphology, RNA and 63 protein leading to the inactivation of SARS-CoV-2 [3,17]. Therefore, it is expected that 64 photocatalysts can inactivate SARS-CoV-2, regardless of the rapidly evolving variants. 65

On the other hand, photocatalysts have three limitations: First, the photocatalytic re-66 action occurs only on the surface of the photocatalyst. Therefore, it is necessary to coat all 67 the surfaces to avoid contamination, or to use it together with a circulator as air purifier. 68 Second, the wavelength of light that can be used to excite the photocatalysts is limited. 69 The wide bandgap (larger than 3 eV) of  $TiO_2$  which is the most common photocatalyst 70 limits the wavelength of the excitation light to the UV region [18]. Thus, narrowing the 71 band gap of TiO<sub>2</sub> is very important for using the TiO<sub>2</sub> photocatalyst under visible light 72 [19]. For example, mixing TiO<sub>2</sub> with silicane (SiH) narrows the band gap (2.082eV) and it 73 can be excited with visible light [20]. Third, since light is necessary for the excitation of 74 photocatalysts, the photocatalytic reaction does not occur in dark conditions, such as 75 while sleeping. 76

In this study, to overcome these limitations, we used a WO<sub>3</sub> photocatalyst containing 77 copper that can applied to a surface by spraying. WO<sub>3</sub> coating kept the surface clear of 78 viral contamination. In addition, unlike TiO<sub>2</sub>, WO<sub>3</sub> could be excited by room light even 79 without mixing with any other compounds such as SiH. Therefore, a light source with 80 specific wavelength was not required. Furthermore, mixing copper with WO<sub>3</sub> particles is 81 expected enable the photocatalyst to inactivate the virus even in the dark, due to the effect 82 of copper. However, there is only one study that reported the inactivation of the SARS-83 CoV-2 Pango lineage A by WO<sub>3</sub> photocatalyst [17]. In this study, we investigated the 84 SARS-CoV-2 inactivation ability of WO<sub>3</sub> photocatalyst both in white light (irradiated by 85 an light emitting diode (LED)) and in darkness, confirmed the time- and concentration-86 dependency of SARS-CoV-2 inactivation by WO<sub>3</sub> photocatalyst, and analyzed the effec-87 tiveness of this photocatalyst against different variants of SARS-CoV-2, according to Jap-88 anese Industrial Standards (JIS). 89

#### 2. Results

#### 2.1. Inactivation of SARS-CoV-2 WK-521 strain by WO<sub>3</sub> photocatalyst.

First, to confirm the inactivation ability of WO<sub>3</sub> photocatalyst against SARS-CoV-2, 92 the WK-521 strain was placed on the WO<sub>3</sub> coated glass and irradiated with 1000 lx light 93 (Figure 1A). As shown in Figure 1B, the titer of SARS-CoV-2 WK-521 strain placed on WO<sub>3</sub> 94 coated glass significantly decreased after irradiation with light for 240 min, compared to 95 before irradiation. In addition, the infectivity of SARS-CoV-2 WK-521 strain placed on 96 WO<sub>3</sub> coated glass in a dark place also decreased. However, this decrease was not to the 97

extent observed in the illuminated sample. Indeed, the mean antiviral activity values were 98 3.04 and 1.50 in with and without light conditions respectively (Figure 1C). This decre-99 ment in the dark condition might be due to the antiviral effect of the copper contained in 100 the WO<sub>3</sub> photocatalyst. On the other hand, no decrease in the titer was observed in the 101 samples placed on the glass without WO<sub>3</sub> coating for 240 min, with or without exposure 102 to light. These results showed that the excitation light itself had no antiviral effect, and the 103 decrease in the titer was due to the effect of the LED-WO<sub>3</sub> photocatalytic reaction in light, 104 and due to the antiviral effect of copper, in darkness. 105

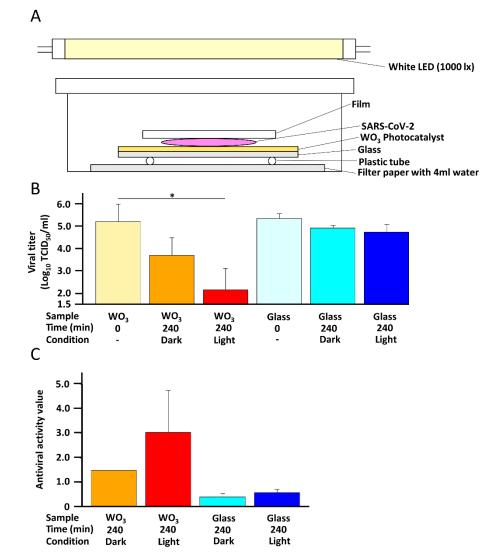


Figure 1. Inactivation of SARS-CoV-2 WK-521 strain by WO<sub>3</sub> photocatalyst. (A) A wet filter paper 107 was placed in a 10 cm dish to avoid dryness. Glass with or without a coating of WO<sub>3</sub> photocatalyst 108 containing copper (100 mg) was placed on a plastic tube which was in turn placed on the filter paper 109 to avoid direct contact with the filter paper. SARS-CoV-2 WK-521 strain (150  $\mu$ L) with a titer of 110  $1 \times 10^{6}$  50% tissue culture infective dose (TCID<sub>50</sub>)/mL was placed on the coated or uncoated glass. 111 WO3 photocatalyst was excited by white LED light with 1000 lx for 0 or 240 min. To confirm the 112 effect of WO3 photocatalyst containing copper in dark condition, SARS-CoV-2 WK-521 strain was 113 placed on the glass with or without a coating of the photocatalyst, and kept in dark for 240 min. (B) 114 Titers of SARS-CoV-2 WK-521 strain were measured using the TCID<sub>50</sub> assay with Vero E6/TMPRSS2 115 cells. Assays were performed in at least 6 wells and the values represent the mean ± standard devi-116 ation (SD) of two independent experiments. Statistical comparisons were performed using Student's 117 t-test. Asterisk indicates a statistically significant difference (\* p < 0.05). (C) Antiviral activity value 118 was calculated using the formula: (The log10 titer of SARS-CoV-2 WK-521 strain of 240min sample) 119 - (The log10 titer of SARS-CoV-2 WK-521 strain of 0 min sample of same sample). 120

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## *2.2. Time- and dose-dependency of the antiviral effects of* WO<sub>3</sub> *photocatalysts.*

Next, to confirm the time-dependence of the antiviral effect of the WO<sub>3</sub> photocatalyst, 122 SARS-CoV-2 WK-521 strain was placed on WO<sub>3</sub> coated glass and irradiated with light for 123 0, 60, 120 and 240 min. As shown in Figure 2A, the viral titer decreased in a time-depend-124 ent manner, and the mean antiviral activity values were 0.66, 1.08 and 2.25 for 60-, 120-125 and 240-min light exposures respectively (Figure 2B). In addition, to confirm the antiviral 126 effect of the WO3 photocatalyst, SARS-CoV-2 WK-521 strain was placed on glasses coated 127 with 10, 30 or 100 mg of WO<sub>3</sub> and irradiated with LED light for 0 or 240 min. As shown in 128 Figure 2C, in the group irradiated with light for 240 min, a decrease in the titer was ob-129 served in all concentrations of WO<sub>3</sub> photocatalyst, compared to the group not exposed to 130 light (0 min), which was significant in the 30 and 100 mg coatings. The antiviral activity 131 values were 2.00, 1.25 and 0.83 for 100, 30 and 10 mg of WO<sub>3</sub> respectively, indicating that 132 the titer of SARS-CoV-2 WK-521 strain was decreased by WO<sub>3</sub> photocatalyst in a dose-133 dependent manner (Figure 2D). In contrast, there was no difference in viral titers among 134 various concentrations on WO3 coated glass in the group which was not irradiated by LED 135 light (0 min, Figure 2C). Our results demonstrated that photocatalysis is the mechanism 136 of inactivation of SARS-CoV-2 WK-521 strain by WO<sub>3</sub>, photocatalytic inactivation of 137 SARS-CoV-2 WK-521 strain by WO3 was dose and time dependent. 138

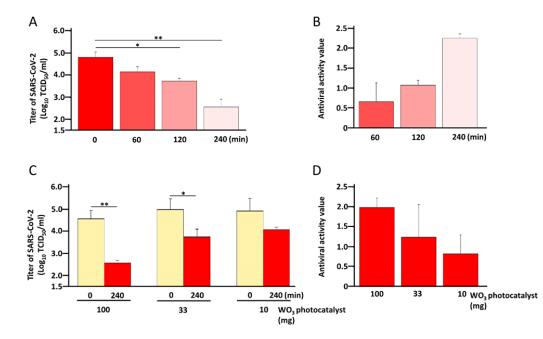


Figure 2. Time- and dose-dependency of the antiviral effects of WO<sub>3</sub> photocatalysts. (A) To confirm 140 time-dependency, SARS-CoV-2 WK-521 strain (150  $\mu$ L) with a titer of 1 × 10<sup>6</sup> 50% tissue culture in-141 fective dose (TCID50)/mL, were placed on glass coated with WO3 photocatalyst (100 mg). WO3 pho-142 tocatalyst was excited by white LED light with 1000 lx for 0, 60, 120 or 240 min. The titer of SARS-143 CoV-2 WK-521 strain was measured using the TCID50 assay with Vero E6/TMPRSS2 cells. Assays 144 were performed in at least 6 wells and the values represent the mean ± standard deviation (SD) of 145 two independent experiments. (B) Antiviral activity value was calculated using the formula: (The 146 log10 titer of SARS-CoV-2 of each time point sample) - (The log10 titer of SARS-CoV-2 of 0min sample 147 of same sample). (C) To confirm concentration-dependency, SARS-CoV-2 WK-521 strain (150 µL) 148 with a titer of 1 × 106 TCID<sub>50</sub>/mL were placed on glass coated with 10, 30 or 100 mg WO<sub>3</sub> photocata-149 lyst. WO<sub>3</sub> photocatalyst was excited by white LED light with 1000 lx for 0 or 240 min. Titers of SARS-150 CoV-2 were measured using the TCID<sup>50</sup> assay with Vero E6/TMPRSS2 cells. Assays were performed 151 in at least 6 wells and the values represent the mean ±SD of two independent experiments. (D) An-152 tiviral activity value was calculated using the formula: (The log10 titer of SARS-CoV-2 WK-521 strain 153 of 240min sample) - (The log10 titer of SARS-CoV-2 WK-521 strain of 0min sample of same concen-154 tration sample). Statistical comparisons were performed using Student's t-test. Asterisk indicates a 155 statistically significant difference (\* p < 0.05; \*\* p < 0.01). 156

Finally, we clarified whether the WO3 photocatalyst exerted antiviral effect against 158 SARS-CoV-2 Omicron variant BA. 2. The Omicron variant BA. 2 was placed on a WO<sub>3</sub> 159 coated glass (150  $\mu$ L with a titer of 1 × 10<sup>7</sup> 50% tissue culture infective dose (TCID<sub>50</sub>)/mL 160 and irradiated with LED light for 0 to 240 min. As shown in Figure 3A, exposure to light 161 for 240 min reduced the titer of this variant, similar to what was observed in the WK-521 162 strain. In addition, the mean antiviral activity after 240 min of photocatalytic reaction on 163 WO<sub>3</sub> was 3.17, which was comparable to that observed in the WK-521 strain (Figure 3B). 164 This result indicated that the WO<sub>3</sub> photocatalyst inactivates SARS-CoV-2 regardless of the 165 variant. 166

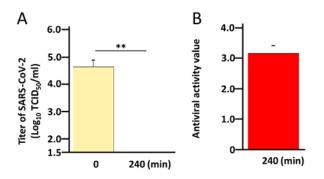


Figure 3. WO3 photocatalysts inactivates SARS-CoV-2 Omicron variant BA. 2. (A) SARS-CoV-2 168 Omicron variant BA. 2 (150  $\mu$ L) with a titer of 1 × 10<sup>7</sup> 50% tissue culture infective dose (TCID<sub>50</sub>)/mL 169 was placed on glass coated with WO<sub>3</sub> photocatalyst (100 mg). WO<sub>3</sub> photocatalyst was excited using 170 a white LED light with 1000 lx for 0 or 240 min. The titer of SARS-CoV-2 was measured using the 171 TCID50 assay with Vero E6/TMPRSS2 cells. Assays were performed in at least 6 wells and the values 172 represent the mean ± standard deviation (SD) of two independent experiments. (B) Antiviral activity 173 value was calculated using the formula: (The log10 titer of SARS-CoV-2 of 240 min sample) - (The 174log10 titer of SARS-CoV-2 of 0 min sample). Statistical comparisons were performed using Student's 175 t-test. Asterisk indicates a statistically significant difference (\*\* p < 0.01). 176

#### 3. Discussion

In this study, we demonstrated that the WO<sub>3</sub> photocatalyst containing copper effec-178 tively inactivated SARS-CoV-2. Indeed, our results provide evidence that WO<sub>3</sub> photocata-179 lytic reaction for 240 min significantly decreased the infectivity of SARS-CoV-2 WK-521 180strain. Additionally, the copper present in the photocatalyst enabled it to inactivate the 181 virus even in darkness. Furthermore, the WO3 photocatalyst containing copper decreased 182 the SARS-CoV-2 WK-521 strain titers in a time-and dose-dependent manner, confirming 183 the photocatalysis induced inactivation of the virus. Our results are supported by a pre-184 vious report that showed the effective inactivation of SARS-CoV-2 by a WO<sub>3</sub> – based vis-185 ible light-responsive photocatalyst on under different temperatures and exposure dura-186 tions [17]. Notably, we demonstrated the effectiveness of a 240 min photocatalytic reaction 187 involving WO<sub>3</sub> photocatalyst containing copper, not only against the SARS-CoV-2 WK-188 521 strain but also against the Omicron variant BA. 2, as indicated by the decreased viral 189 titers. The present study is the first to report that a WO<sub>3</sub> photocatalytic reaction can inac-190 tivate SARS-CoV-2, regardless of the variant. 191

It is previously reported that the mechanisms involved in the inactivation of SARS-192 CoV-2 by photocatalysis are damage of viral morphology, RNA, and protein [3,17]. In this 193 study, the inactivation of the SARS-CoV-2 WK-521 strain and the Omicron variant BA. 2 194 by WO<sub>3</sub> photocatalyst containing copper was demonstrated. This suggests that even if the 195 virus is mutated, a photocatalytic reaction by WO<sub>3</sub> can achieve viral inactivation by dam-196 aging the viral protein, RNA and lipid bilayer, irrespective of the variant. Hence, this WO3 197 photocatalyst containing copper, could be effective against the variants of SARS-CoV-2 198 which may in the future. 199

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WO<sub>3</sub> photocatalyst containing copper inactivated SARS-CoV-2 not only upon irradi-200 ation with light, but also in dark condition as well. It has been reported previously that 201 copper nanoparticles can inactivate SARS-CoV-2 [21]. In addition, copper oxide (CuO) 202 nanoclusters grafted with titanium dioxide also inactivated SARS-CoV-2 alfa, beta, 203 gamma and delta variants under illumination and in dark conditions as well [22]. These 204 observations suggest that copper is responsible for the inactivation SARS-CoV-2 WK-521 205 strain under dark condition, observed in this study, indicating that the WO<sub>3</sub> photocatalyst 206 containing copper can be effective even in the night time. 207

Since WO<sub>3</sub> photocatalyst containing copper was excited by a white LED in this study, 208 it is evident that this photocatalyst works effectively in indoor environment, without the 209 necessity for any specific light source. In addition, unlike UV, WO<sub>3</sub> photocatalyst is harm-210 less to the human body. Therefore, it is expected that WO<sub>3</sub> photocatalyst containing cop-211 per can be used for disinfection on surfaces which are touched regularly by many people, 212 thereby preventing the viral spread. Thus, this study demonstrated that WO<sub>3</sub> photocata-213 lyst containing copper could be effectively applied to control SARS-CoV-2 transmission 214 and mitigate the ongoing COVID-19 pandemic. 215

### 4. Materials and Methods

## 4.1. Virus and cell culture

Vero E6 cells which express the transmembrane serine protease TMPRSS2 (Vero 218 E6/TMPRSS2 (JCRB1819)) were maintained in Dulbecco's Modified Eagle's Medium 219 (DMEM, Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% heat-in-220 activated fetal bovine serum (FBS, Thermo Fisher Scientific) at 37 °C with 5% CO<sub>2</sub>. The 221 WK-521 strain (Pango lineage A; 2019-nCoV/Japan/TY/WK-521/2020) [23] and Omicron 222 variant BA. 2 (hCoV-19/JPN/TY40-385/2022 strain) of SARS-CoV-2 were cultured and 223 quantified using Vero E6/TMPRSS2 cells in DMEM containing 2% FBS. The infectivity of 224 the viral strains was calculated by titrating in Vero E6/TMPRSS2 cells using the TCID50 225 assay and applying the Reed-Muench method [24]. 226

## 4.2. Inactivation of SARS-CoV-2 by WO3 Photocatalytic Reaction

The photocatalytic reaction was performed according to JIS R1752:2020 [25] with a 228 minor modification (Figure 1A). Briefly, filter paper was placed at the bottom of the 10 cm 229 dish and got wet by 4 mL sterilized water for moisture preservation. To avoid directly 230 touching the filter paper, a plastic tube was placed on the filter paper and a glass coated 231 with 100 mg WO<sub>3</sub> photocatalyst containing copper (NFE2-W; Chemical Technology Co., 232 Ltd., Takaishi, Osaka, Japan) was placed on top of the plastic tube. 150 µL of the WK-521 233 strain with a titer of  $1 \times 10^6$  TCID<sub>50</sub>/mL or the Omicron variant BA. 2 with a titer of  $1 \times 10^7$ 234 TCID<sub>50</sub>/mL was placed on the WO<sub>3</sub> coated glass and spread by covering it with a film. 235 Glass without WO<sub>3</sub> coating was used as a negative control. The samples were then illumi-236 nated with 1000 lx light using a white LED (BBZ T13 Silver; Dongguan oushi Electronic 237 Technology Co., Ltd, Dongguan, Guangdong China) for 240 min, or not illuminated. After 238 illumination, the samples were washed by immersing in 8 mL phosphate buffered saline 239 (PBS). As time control, the virus was immediately collected after placing the droplet on 240 glass with or without  $WO_3$  coating (0 min). To confirm the time-dependency, the SARS-241 CoV-2 WK-521 strain was placed on the 100 mg WO<sub>3</sub> coated glass and illuminated for 0, 242 60,120 and 240 min. To observe the concentration-dependency, the SARS-CoV-2 WK-521 243 strain was placed on glass coated with 100, 30 or 10mg WO<sub>3</sub> and illuminated for 0- and 244 240-min. SARS-CoV-2 titers in all experiments was measured by TCID<sub>50</sub> assay. 245 246

The photocatalytic inactivation efficiency was defined as follows:

Antiviral activity value =  $\log_{10} (N_t) - \log_{10} (N_0)$ 

where  $N_i$  represents the virus titer of the photocatalytically treated specimens after 248 irradiation for t hours;  $N_0$  represents the virus titer of the photocatalytically treated spec-249 imens just after inoculation (0 min). 250

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	4.3. Statistical analysis	251
	Statistical comparisons were performed using Student's t-test. $p$ values < 0.05 were considered statistically significant.	252 253
	5. Conclusions	254
	This is the first report showing that WO3 photocatalyst inactivates Omicron variant	255
	BA. 2 as well as SARS-CoV-2 WK-521 strain, indicating the effectiveness of this photocata-	256
	lyst against the virus, regardless of the variant. In addition, WO <sub>3</sub> photocatalyst containing	257
	copper can inactivate the virus using a simple white light or even in dark conditions, indi-	258
	cating its potential for wide application. In conclusion, WO <sub>3</sub> photocatalyst containing cop-	259
	per could be a very effective tool for controlling the spread of SARS-CoV-2.	260
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	performed the experiments: K.M, R.M., and Y.A. Analyzed the data: R.M., and Y.A. Supervised this	262
	experiment: Y.A., K.M, K.H, Y.M. (Yosuke Mori) and Y.M. (Yasunobu Matsumoto). Contributed	263
	reagents/materials/analysis tools: Y.A., K.M., K.H, Y.M. (Yosuke Mori), and T.K. Wrote the paper: R.M., and Y.A. All authors have read and agreed to the published version of the manuscript.	264 265
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	Conflicts of Interest: The authors declare no conflict of interest.	272
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